

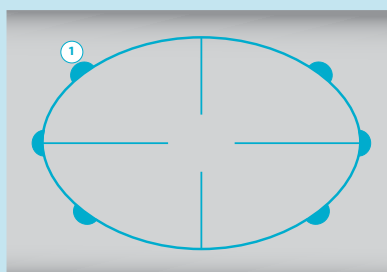
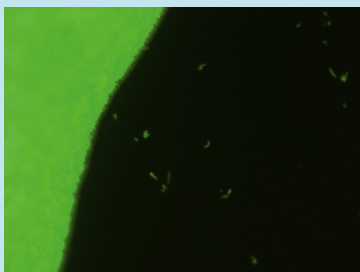
Patient ID Area: Etching area for proper labeling of specimen or patient ID

Fluorescent Smear Guide: A thin fluorescent oval that provides users with a standard area for smear preparation

Starter Circles: Large fluorescent markers that provide a reference point for focusing on slide

Crosshairs: Fluorescent lines that aid the user in quantifying fields as well as allowing proper focusing throughout the examination pathway, as explained below

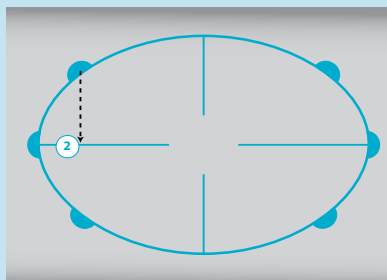
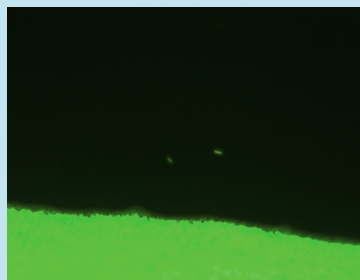
1. Upper Left Starter Circle



Focus on this upper left starter circle, and travel down toward the left crosshair. AFB may gather close to the fluorescing mark, as shown in the image to the left.

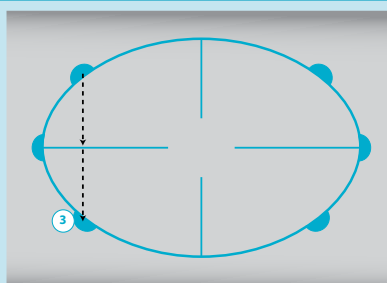
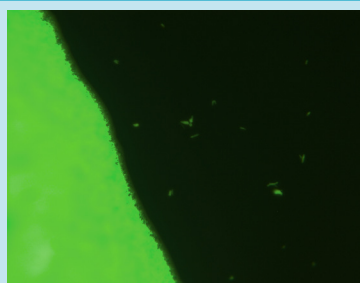
Note: All photographs in this section were taken at 600x magnification with limited digital modification.

2. Left Crosshair



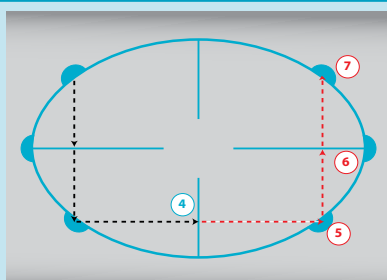
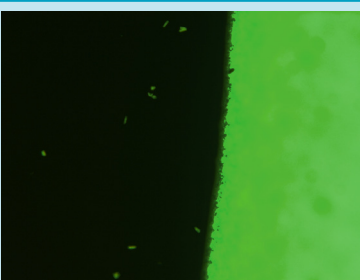
Once the crosshair has been reached, refocus (if necessary) and continue downward to the bottom-left starter circle. Two (2) fluorescing bacilli are seen in this picture.

3. Lower Left Starter Circle



After reaching the lower left starter circle, refocus (if necessary), and continue moving right toward the bottom crosshair. Both single and clumped bacilli are shown in the image to the left.

4. Bottom Crosshair



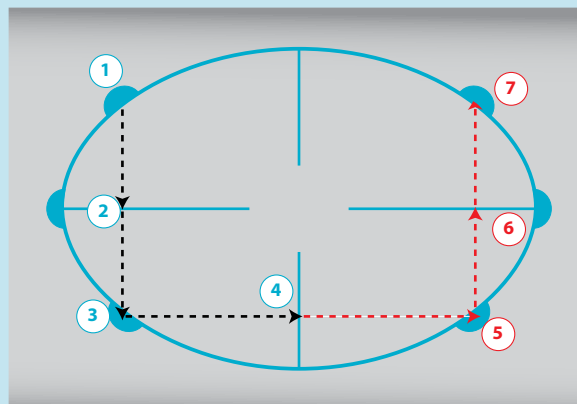
Once the final crosshair has been reached, quantify the number (#) of bacilli seen, and use the standard recording system on the next page to report findings, based on magnification.

If further fields are needed for viewing (to confirm a scanty or an unclear result), observe the sample along the path designated by numbers 4-7. Use the same methods as previously described.

SureFocus™ Microscope Slide User Guide

Calculating Fields Viewed

By moving along the dotted lines from point 1 to point 4 (as seen to the right) the reader will view an appropriate number of fields to quantify AFB based on the WHO standards for AFB Smear Grading (as seen below). Use the following chart to determine the approximate number of fields read:



Total Magnification	Approximate # of Fields Viewed (from points 1-4)
200x	26
400x	52
600x	78

Reporting Results

Follow this recommended grading scale to quantify AFB (based on the WHO guidelines for AFB smear grading¹):

Ziehl-Neelsen Stain	Fluorescence Microscopy: # of AFB and Fields Observed Using SureFocus Slide (approximate, using points 1-4)			Report
	1000x (~100 fields)	200x (~26 fields)	400x (~52 fields)	
0	0	0	0	No AFB Seen
1-9 AFB total	1-29 AFB total	1-19 AFB total	1-12 AFB total	Report Exact Count
10-99 AFB total	1-10 AFB per field	20-199 AFB total	13-239 AFB total	+
1-10 per field	10-100 AFB per field	5-50 AFB per field	3-30 AFB per field	++
>10 per field	>100 AFB per field	>50 AFB per field	>30 AFB per field	+++

1. Fluorescent Stain Preparation. WHO laboratory services in tuberculosis control. Part II: Microscopy. WHO/TB/98.258. Geneva, Switzerland: WHO, 1998: 31-34.)

Slide Reading Tips



1. Use positive and negative control slides as a reference and quality control for each batch of slides stained. These controls also aid the viewer in reviewing the morphological characteristics of fluorescing organisms.
2. Bacilli size is roughly 1-10 μm long, and may appear as curved or bent rods. Some non-tuberculous species of mycobacteria may appear more coccoid or have longer rods.
3. Bacilli may appear alone or in small groups. Large clumps of organisms are rarely seen from direct specimen smears.
4. Fluorescing bacilli should appear as yellow-green rods against a darker background.
5. Non-mycobacterial artifacts are sometimes present in specimens and may fluoresce. Examples include hair, fiber and food particles. To avoid confusion, it is important to learn what a fluorescing bacillus looks like.